## **BIELSCHOWSKY'S METHOD FOR NEUROFIBRILS (Microwave)**

PURPOSE:	For In Vitro Diagnostic Use: Intended for the qualitative demonstration of senile plaques and fibrillary tangles.
PRINCIPLE:	Nerve fibers are first sensitized with a Silver Solution and then they absorb silver from the Diamine Silver Solution. The silver is reduced to metallic silver by Formalin and the excess is removed by Thiosulfate.
CONTROL:	Cerebral cortex of a known case of Alzheimer's Control Slides can be purchased from Histology Control Systems. See inside back cover, Item# cs031.
SPECIMEN PREPARATION:	15% Buffered Formalin, for at least 2 weeks, fixed, paraffin embedded sections cut at 8 micrometers
SOLUTIONS:	<ol> <li>Silver Nitrate 1% Aqueous Item# s1888</li> <li>Ammonium Hydroxide Concentrated Item# c804</li> <li>Silver Nitrate 5% Aqueous Item# s1890</li> <li>Ammonium Hydroxide 1% Solution Item# s2338</li> <li>Sodium Thiosulfate 2% Aqueous Item# s280</li> <li>Bielschowsky's Developing Solution Item# s2788</li> <li>Solutions can be purchased separately from Poly Scientific.</li> </ol>
NOTES:	Use acid cleaned glassware. Do not crowd slides in the staining container. Use the same silver solution throughout the technique. Variations in timing may occur due to the power wattage of the microwave oven. Provided times and power levels are based on 1000 watt microwave oven.
REFERENCE:	Luna, Lee G. <u>Histopathologic Methods and Color Atlas of Special</u> Stains and Tissue Artifacts. American Histolabs Inc. Gaithersburg, MD. 1992. p. 484.

## STAINING PROCEDURE:

- 1. Deparaffinize and hydrate slides to distilled water.
- 2. Place slides in 40 mL of Silver Nitrate 1% Aqueous in a plastic coplin jar. Microwave at power level 3 (180 W) for 1 minute. Dip slides several times in warm solution and let set for 15 minutes.
- 3. Rinse slides in distilled water.
- 4. To make Ammoniacal Silver Nitrate, pour the warm Silver Nitrate 1% from step 2 into a 125 mL flask. Add Ammonium Hydroxide Concentrated drop by drop, constantly shaking, until the solution clears. Next add Silver Nitrate 5%, drop by drop, shaking constantly until the solution becomes slightly cloudy.
- 5. Pour Ammoniacal Silver Nitrate Solution prepared in step 4 into a plastic coplin jar. Add slides and microwave at power level 3 (180 W) for 1 minute. Dip slides up and down in the warm solution and let set for 15 minutes.
- 6. Place slides directly into Ammonium Hydroxide 1% Solution for not more than 20 seconds.
- 7. Add 3 drops of the Developer into the Ammoniacal Silver Nitrate Solution from step 5 and mix with glass rod. Place slides immediately into the solution for 2-3 minutes or until the sections turn brown. *Note:* The solution will turn a gray color and a sheen of silver will form on the sides of the coplin jar and sometimes on the glass slides but not on the tissue sections.
- 8. Place slides in Ammonium Hydroxide 1% Solution for no longer than 15 seconds.
- 9. Rinse slides in 3 changes of distilled water.
- 10. Carefully wipe off silver from both sides of slides. Do not touch tissue section.
- 11. Place slides in Sodium Thiosulfate 2% Aqueous for 30 seconds.
- 12. Rinse slides in 4 changes of distilled water.
- 13. Dehydrate in 95% Alcohol, Absolute Alcohol and clear in Xylene, 2 changes each.
- 14. Mount with Poly Mount (Item# s2153) or any other acceptable mounting medium.

## **RESULTS**:

Neurofibrillary Tangles and Neuritic Plaques of Alzheimer's Disease	Dark Brown to Black
Neuromelanin, Lipofuscin	Densely Black & Coarsely Granular
Axons	Brown to Black
Amyloid	Dense Black Cores
Background	Yellow-Brown

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