

GIMENEZ METHOD FOR HELICOBACTER PYLORI

PURPOSE:	For In Vitro Diagnostic Use: Intended for the qualitative demonstration of Helicobacter Pylori in tissue.
PRINCIPLE:	The staining solution is able to dissolve into the lipid capsule of the bacteria and then resist being destained by the water rinsing. The counterstain is for contrast.
CONTROL:	Any tissue known positive for Helicobacter Pylori <i>Control Slides can be purchased from Histology Control Systems. See inside back cover, Item# cs033.</i>
SPECIMEN PREPARATION:	Formalin fixed, paraffin embedded sections cut at 3-5 micrometers
SOLUTIONS:	1. Carbol Fuchsin Ziehl Neelsen Item# s162 2. Phosphate Buffer 0.1M pH 7.5 Item# s2078 <u>Working Carbol Fuchsin Solution:</u> Carbol Fuchsin Ziehl Neelsen.....15.2 mL Phosphate Buffer 0.1M pH 7.5.....38.0 mL A precipitate will form immediately. Mix well and filter. Filter again immediately before use. Working Solution is stable for 48 hours. 3. Malachite Green 0.8% Aqueous Item# s2198 <i>Solutions can be purchased separately from Poly Scientific.</i>
NOTES:	
REFERENCE:	Luna, Lee G. <u>Histopathologic Methods and Color Atlas of Special Stains and Tissue Artifacts</u> . American Histolabs Inc. Gaithersburg, MD. 1992. pp. 219-220.

STAINING PROCEDURE:

1. Deparaffinize and hydrate slides to distilled water.
2. Working Carbol Fuchsin Solution for 2-5 minutes.
3. Wash thoroughly in water.
4. Malachite Green 0.8% Aqueous for 15–20 seconds.
5. Wash thoroughly in water.
6. Repeat steps 4 and 5 until sections become blue-green.
7. Blot and allow sections to air dry.
8. Clear in Xylene and mount with Poly Mount (Item# s2153) or any other acceptable mounting medium.

RESULTS:

Helicobacter Pylori, Rickettsia and Other Bacteria.....Bright Red
Nuclei.....Blue-Green
Background Light-Green

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Revision: B-18

